## DNA POLYMORPHISMS IN BOREAL OWLS (AEGOLIUS FUNEREUS)

JANNE BEHEIM

Department of Animal Science, Agricultural University of Norway, P.O. Box 5025, 1432 Ås-NLH, Norway

Katrine Eldegard

Department of Biology and Nature Conservation, Agricultural University of Norway, P.O. Box 5014, 1432 Ås-NLH, Norway

GRO BJØRNSTAD

Department of Morphology, Genetics and Aquatic Biology, Norwegian College of Veterinary Medicine, P.O. Box 8146 Dep., 0033 Oslo, Norway

MATS ISAKSSON

Department of Genetics, Uppsala University, Box 7003, 750 07 Uppsala, Sweden

GEIR SONERUD Department of Biology and Nature Conservation, Agricultural University of Norway, P.O. Box 5014, 1432 Ås-NLH, Norway

> OLAV HEIE Drøbak Medical Center, Storgaten 18, 1440 Drøbak, Norway

> > Helge Klungland<sup>1</sup>

Department of Animal Science, Agricultural University of Norway, P.O. Box 5025, 1432 Ås-NLH, Norway

KEY WORDS: Boreal Owl; Tengmalm's Owl; Aegolius funereus; microsatellite, DNA polymorphisms; variability.

Molecular analyses of polymorphic DNA-fragments are widely used in phylogenetic studies to recognize individuals, to evaluate mating strategies, and to study genetic diversity (Lawless et al. 1997, Primmer and Ellegren 1998). A limiting factor in studies that depend on species-specific variation is the number of available markers. Due to the conservational nature of DNA across species, polymorphic regions that are localized in one species will often be of great use in a number of related species. This is also the case for microsatellites, which are often localized in less conserved areas (Chambers and MacAvoy 2000). The main focus of this work was to establish DNA polymorphism in the Boreal Owl (Aegolius funereus funereus) that would be useful for testing paternity, inbreeding, and population genetics. Microsatellites, characterized by short, tandemly-repeated, and highlypolymorphic sequences, were chosen for the analysis. These markers have previously been used for cross-species amplification in birds (Primmer et al. 1996), and in several other species. Although microsatellites are highly polymorphic (varying number of tandemly-repeated motifs), sequences flanking the microsatellite are still conserved enough to be present across related species, and are used for primer binding. As expected, a negative relationship between microsatellite performance and evolutionary distance has been observed (Primmer et al. 1996).

## Methods

Blood samples were collected from 44 unrelated freeranging adult Boreal Owls (Tengmalm's Owl) nesting in Hedmark County, Norway (ca. 61°N, 11°E) in 1998. Natal as well as female breeding dispersal is extensive in the Boreal Owl, causing genetic swamping over large areas (Sonerud et al. 1988). DNA was isolated following standard protocols (Seutin et al. 1991, Krokene et al. 1996) Amplification of microsatellites in Boreal Owl (Table 1) was based on sequences obtained from the Eurasian Eagle-Owl (Bubo bubo; Isaksson and Tegelström 2002) Among the microsatellites used in this study Bb111 and Bb126 are GA repeats, whereas, the remaining satellites are CA repeats. Reactions were carried out in 10 µl containing 50 ng of genomic DNA, 0.5 U Taq polymerase, enclosed buffer (Perkin Elmer), 2.5 pmol of each primer and 0.2 mM of each dNTP. Genomic DNA was denatured for 3 min at 94°C prior to amplification. The polymerase chain reaction (PCR-amplification) was run for 35 cycles at 94°C (denaturation) for 15 sec, annealing for 15 sec, and elongation at 72°C for 30 sec. Annealing temperatures varied from 45°C (Bb42) to 48°C (Bb100, Bb101,

<sup>&</sup>lt;sup>1</sup> Corresponding author's present address: Department of Laboratory Medicine, Faculty of Medicine, Norwegian University of Science and Technology, St. Olavus Hospital, Morfologibygget, 7006 Trondheim, Norway; e-mail address: helge.klungland@medisin.ntnu.no

Marker	PRIMER SEQUENCES		
Bb20 F	GTGGTGGCACGGCTTGT		
Bb20 R	TGTCAAGAGGAAGCATAAAATACAT		
<b>B</b> b120 F	TAATGGTGCTGCTGGTGGAAG		
Bb120 R	CATGTGTAGGTGTGGGAGAGAA		
Bb134 F	TTTCTCCACGCTTCCTTTTCATA		
Bb134 R	AGAAGAATGGCTGGCAAGACTC		

and Bb145) to 50°C (Bb111 and Bb134) and 52°C (Bb126). Successful amplification of Bb20, Bb120, and Bb131 was not obtained at any annealing temperatures. Microsatellites were analyzed on an ABI 373 sequencer.

## **RESULTS AND DISCUSSION**

Of the ten primer pairs characterized in eagle-owls, seven successfully amplified DNA from Boreal Owl (Table 2). Five of these were polymorphic in Boreal Owl, where-

Table 2. Length of alleles, allele frequencies and heterozygosity among 44 unrelated Boreal Owls for seven microsatellites. Microsatellite markers Bb20, Bb120 and Bb131 did not amplify DNA successfully from Boreal Owl.

Micro- satellite Marker	Allele Lengths	Allele Frequencies	Observed Hetero- zygosity
Bb42	304 bp	1.000	0
Bb100	296 bp 298 bp	$0.761 \\ 0.239$	0.30
Bb101	185 bp 187 bp 189 bp 191 bp	0.477 0.034 0.034 0.455	0.57
Bb111	201 bp 203 bp 205 bp 207 bp 209 bp 211 bp	$\begin{array}{c} 0.023 \\ 0.011 \\ 0.080 \\ 0.625 \\ 0.136 \\ 0.080 \end{array}$	0.61
Bb126	213 bp 185 bp 187 bp	$0.045 \\ 0.989 \\ 0.011$	0.02
Bb134	144 bp	1.000	0
Bb145	242 bp 256 bp	$0.898 \\ 0.102$	0.18

as, the remaining two were monomorphic within the individuals tested in our analysis. Because Boreal Owls and eagle-owls are among the most distantly related species within the Strigidae family (Mindell et al. 1997), these microsatellites may be of potential use in most species within this family. Our findings could therefore be of great importance for the analysis of population genetics, as well as for parental testing in a wide variety of species within the Strigidae family.

RESUMEN.—Hemos utilizado los pares de indicadores con base en secuencias del gran búho euroasiático con el fin de ampliar exitosamente siete microsatélites de loci en el búho boreal (*Aegolius funereus funereus*), de los cuales cinco fueron polimorfos. El número de alelos por locus variaron entre dos a siete. La conservación de los microsatélites de loci entre el búho boreal y el gran búho euroasiático indica que las secuencias del gran búho pueden ser útiles en estudios moleculares para la mayoría de especies de la familia strigidae.

[Traducción de César Márquez]

## LITERATURE CITED

- CHAMBERS, G.K. AND E.S. MACAVOY. 2000. Microsatellites consensus and controversy. *Comp. Biochem. Physiol* 126:455–476.
- ISAKSSON, M. AND H. TEGELSTRÖM. 2002. Isolation and characterization of polymorphic microsatellite markers in a captive population of the eagle-owl, *Bubo bubo*, used for supportive breeding. *Mol. Ecol. Notes* 2:91–93.
- KROKENE, C., K. ANTHONISEN, J.T. LIFJELD, AND T. AMUNDSEN. 1996. Paternity and paternity assurance behaviour in the Bluethroat. *Luscinia s. svecica. Anim. Behav.* 52:405–417.
- LAWLESS, S.H., G. RITCHISON, P.H. KLATT, AND D.F. WEST-NEAT. 1997. The mating strategies of Eastern Screech-Owls: a genetic analysis. *Condor* 99:213–217.
- MINDELL, D.P., M.D. SORENSON, C.J. HUDDLESTON, H.C. MI-RANDA, JR., A. KNIGHT, S.J. SAWCHUK, AND T. YURI. 1997 Phylogenetic relationships among and within select avian orders based on mitochondrial DNA. Pages 213–247 *in* D.P. Mindell [ED.], Avian molecular evolution and systematics. Academic Press, London, U.K.
- PRIMMER, C.R. AND H. ELLEGREN. 1998. Patterns of molecular evolution in avian microsatellites. *Mol. Biol. Evol.* 15:997–1008.
- ——, A.P. MOLLER, AND H. ELLEGREN. 1996. A widerange survey of cross-species microsatellite amplification in birds. *Mol. Ecol.* 5:365–378.
- SEUTIN, G., B.N. WHITE, AND P.T. BOAG. 1991. Preservation of avian blood and tissue samples for DNA analysis. *Can. J. Zool.* 69:82–90.
- SONERUD, G.A., R. SOLHEIM, AND K. PRESTRUD. 1988. Dispersal of Tengmalm's Owl *Aegolius funereus* in relation to prey availability and nesting success. *Ornis Scand* 19:175–181.

Received 30 August 2001; accepted 20 April 2002